# Evaluation of Antipsychotic Properties of Aqueous Extract of *Lophira alata* (Ochnaceae) and *Afzelia africana* (Leguminosae) Stem Barks in Rats

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#### ABSTRACT

Stem barks of *Lophira alata* (family: Ochnaceae) and *Afzelia africana* (family: Leguminosae) are used in ethnomedicine for the management of psychosis. Since no existing data was found on their anti-psychotic properties, this study was carried out to evaluate their ability to reduce amphetamine induced stereotypy in rats. The oral mean lethal dose (LD<sub>50</sub>) of both extracts was estimated and preliminary phytochemical screening was conducted. *Lophira alata* and *Afzelia africana* extract (6.25, 12.5 and 25 mg/kg *po*) was investigated for antipsychotic potential on amphetamine induced stereotypy model in rats. The LD<sub>50</sub> of each of the plants was estimated to be greater than 5000 mg/kg. Oral administration of *Lophira alata* and *Afzelia africana* extract produced a significant reduction (p<0.05) in locomotive activity and in episodes of rearing and sniffing typical of stereotypy in rats. The results obtained suggest that the aqueous stem bark extract of *Lophira alata and Afzelia africana* possess antipsychotic properties which may account for their use in ethnomedicine.

Key words: Lophira alata, Afzelia africana, amphetamine, locomotion.

#### INTRODUCTION

Traditional medicine is defined as the sum total of knowledge, skills and practices based on the theories, beliefs and experiences indigenous to different cultures that are used to maintain health, as well as to prevent, diagnose, improve or treat physical and mental illness. It forms the mainstay of treatment for majority of the inhabitants of Africa. In industrialized countries, adaptations of traditional medicine are termed complementary or alternative medicine (WHO, 2003).

Schizophrenia, a form of mental disorder characterized by psychotic symptoms affects about 1% of the population and ranks among the top five causes of disability in adults (Hodgins, 1992; Murray and Lopez, 1996). Drugs currently used for clinical management of psychotic illnesses are dopamine receptor antagonists, some of which also antagonize serotonergic, muscarinic and alpha adrenoceptors. They can be divided into two groups viz :

(i) typical or first generation anti-psychotics such as chlorpromazine, haloperidol, fluphenazine, flupentixol and clopentixol which produce unwanted motor side effects including acute dystonias and tardive dyskinesia (extra-pyramidal side effects).

(ii) atypical or second generation anti-psychotic drugs such as clozapine, risperidone, sertindole, quatiapine, aripiprazole, olazapine and zotepine which are associated with lower incidence of extrapyramidal side effects (Rang *et al.*, 2007; Meltzer, 2009).

Herbal medicines form the cornerstone of therapy for management of psychiatric disorders including psychotic symptoms and schizophrenia in some parts of the world where leaves, stem barks, roots, flowers and fruits of various plants are often used in combination with different rituals for treatment of mental disorders (Ibrahim et al., 2007; Magaji et al., 2008; Romeiras et al., 2012; Ahmed and Azam, 2014). Different parts of Afzelia africana, Annona senegalensis, Datura metel, Euphorbia hirta, Ficus hirta, Ginkgo biloba, Hypericum perforatum, Lophira alata, Milicia excels, Morinda citrifolia, Ocimum americanum, Panax ginseng, Piper retrofractum, Rauwolfia vomitoria, Thevetia peruvian, Valeriana officinalis, Ximenia americana are used in folkloric and in alternative and compl ementary Medicine for management of psychiatric illness. They are used in form of decoctions, juice extracts, taken as food or made into pills for oral consumption (Gill, 1992; Iwu, 1993; Iwu and Wootton, 2002; Ibrahim et al., 2007; Pieme et al., 2008; Sofowora, 2008; Sonibare et al., 2008; Sonibare et al., 2011; Romeiras et al., 2012; Ahmed and Azam, 2014)

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Lophira alata locally known in many parts of Nigeria as red iron wood, Namijin kadai (Hausa), Ekki (Yoruba), Aba, Akufo or ugiri nwautobo (Igbo), is used for its analgesic, anti-inflammatory, anxiolytic, anti-malarial, sedative and antiepileptic effects (Bolza and Keating, 1972; Burkill, 1985; Tih et al., 1994, Ibrahim et al., 2007; Musa et al., 2013; Ajibove et al., 2014; Falade et al., 2014, Balde et al., 2015). Afzelia africana locally known as Lucky bean tree or Kawo (Hausa), Apa (Yoruba) Akpalata (Igbo) is credited with antiepileptic, antipyretic, analgesic, antidiabetic, antimicrobial, antipsychotic and wound healing properties (Burkill, 1985; Adjanohoun et al., 1986; Onweluzo et al., 1995; Balde et al., 2006; Akah et al., 2007; Magassouba et al., 2007; Akinpelu et al., 2008). A decoction of the stem bark of Lophira alata and Afzelia africana are used together in traditional medicine for the management of psychosis and as a sedative agent (Ibrahim et al., 2007).

A search through existing Literature did not reveal any studies on the antipsychotic properties of these plants; this study was therefore carried out to evaluate their activity in animal models of psychosis.

## MATERIALS AND METHODS

### **Plant materials**

Leaves and stem barks of Lophira alata obtained from Idi Mongoro forest, Ore Ondo State-Nigeria in November 2012 were identified by authenticated by Mr. K. A. Adeniji and Mr. O. A. Ugboga of the Forest Research Institute of Ibadan, Oyo State-Nigeria where a voucher specimen (FHI 109820) was deposited for future reference. Leaves and stem bark of Afzelia africana obtained from Okuomu forest in Benin City, Edo State in October 2011 were identified by Mr. S. Nweke of the Department of Pharmacognosy, Faculty of Pharmacy, University of Benin, Benin City, Edo State and authenticated by Mr. K. A. Adeniji and Mr. O. A. Ugboga of the Forest Research Institute of Ibadan, Oyo State-Nigeria where a voucher specimen (FHI 110362) was deposited for future reference The stem barks were cleaned, dried and reduced to powdered form by milling. Three hundred (300) grams of each plant powder was macerated with 2 liters of distilled water for 24 hours, with occasional shaking at room temperature. The extracts were decanted, filtered and concentrated over a water bath at 40°C. Fresh solutions of both extracts in distilled water were prepared for use each day and a mixture of equal amounts of Lophira alata and Afzelia africana was made just before drug administration.

## **Phytochemical Screening**

Phytochemical screening was carried out in accordance with the standard protocol as described

by Trease and Evans (1983). The extracts were screened for the presence of alkaloids, tannins, saponins, flavonoids, glycosides, phytosterols and triterpenoids.

## Animals

Male Wistar rats weighing 180-230 grams procured from the Department of Pharmacology and Toxicology, University of Benin, Benin-City, Edo State, Nigeria were used for the study. They were kept in polypropylene cages at the Animal House Facility of the Department of Pharmacology and Toxicology, University of Benin, Benin City, Edo State, Nigeria. The animals were maintained under standard laboratory conditions with access to animal feed and clean water *ad libitum*. Adequate hygiene was maintained through daily cleaning of cages. Handling of the animals was done according to standard protocols for the use of laboratory animals of the National institute of Health (NIH, 2002).

## Drugs/Chemicals

Amphetamine (Tocris Bioscience McKinley, Minneapolis USA) and haloperidol (Janssen Pharmaceuticals, Lagos Nigeria) were used in this study.

## Acute toxicity study in mice

The acute toxicity and lethality of Lophira alata and Afzelia africana was determined in rats using the method described by Lorke (1983). Briefly, nine rats randomly distributed into three groups (n=3) were orally administered 10, 100 and 1000 mg/kg of Lophira alata respectively and observed for signs of toxicity and death for 24 hours. Based on the outcome of the first phase, 1600, 2900 and 5000 mg/kg Lophira alata were administered to a fresh batch of animals (n=1) and observed for number of deaths for 24 hours. The same procedure was conducted for Afzelia africana extract. The median lethal (LD<sub>50</sub>) dose was estimated as the geometric mean of the lowest lethal dose at which the animal died and the highest non-lethal dose at the animal survived.

## **Behavioural Studies**

Anti-psychotic potential of *Lophira alata* and *Afzelia africana* was evaluated using the amphetamine induced model of stereotypy in rats (Ayoka *et al.*, 2006). Male Wistar rats were brought to the test room one hour before the test in order to acclimatize. Each animal was further introduced into the test box for 30 minutes prior to drug administration. Rats were randomly assigned into groups of 5 each and treated with distilled water orally (group I), 3 dose levels of

Lophira alata and Afzelia africana orally (6.25, 12.5 and 25 mg/kg, groups II-IV respectively) and haloperidol 2 mg/kg *i.p* (group V) followed thirty minutes later by 2 mg/kg amphetamine administered subcutaneously (s.c). One hour post administration of amphetamine, animals were placed individually in the open field box measuring 40×20×18 cm and several indices were scored according to the method described by Costall and Naylor with some modifications (Costall and Naylor, 1973). Locomotive activity indicated by number of lines crossed, stereotypy measured by episodes and frequency of rearing and sniffing were scored by observers blinded to treatment in blocks of 5 minutes for one hour (Costall and Naylor, 1973). The open field box was cleaned with 70% alcohol after each animal was removed. In order to simulate psychosis which is often associated with hyperdoparminergic activity in the nigrostriatal and mesolimbic pathways (Burt et al., 1977; Abi-Dargham et al., 2005), a separate set of animals (5 per group) were pretreated with amphetamine 2 mg/kg s.c followed thirty minutes later by distilled water orally (group I), 6.25, 12.5 and 25 mg/kg, of Lophira alata and Afzelia africana orally (groups II-IV respectively) and haloperidol 2 mg/kg *i.p* (group V). One hour later indices of locomotive activity and stereotypy were scored by unbiased observers as previously described (Costall and Naylor, 1973).

#### **Statistical Analysis**

Data are expressed as mean  $\pm$  SEM and statistically analyzed by one-way ANOVA followed by the Tukey test. A probability value of p <0.05 was considered statistically significant. All statistical analyses were performed using Sigma Plot version 11.0 for Windows.

## RESULTS

Percentage yield post extraction for *Lophira alata* and *Afzelia africana* was 10% and 4% respectively.

The preliminary phytochemical screening of aqueous extract of *Lophira alata* and *Afzelia africana* stem barks revealed the presence of alkaloids, tannins, saponins, flavonoids, triterpenoids, phytosterols and glycosides (Table 1).

The oral  $LD_{50}$  of *Lophira alata* and *Afzelia africana* in rats was estimated to be greater than 5000 mg/kg (for each plant).

Pretreatment with *Lophira alata* and *Afzelia africana* decreased locomotive activity at all 3 dose levels in the first 20 minutes of observation; there was a significant difference (p<0.05) between test groups and amphetamine treated groups within this time frame. Locomotive activity increased in the last forty minutes of observation in the *Lophira alata and Afzelia africana* treated groups, locomotion here was not significantly different from amphetamine treated groups (Figure 1).

Post treatment with *Lophira alata* and *Afzelia africana* resulted in reduced locomotive activity with 12.5 mg dose level for the entire 1 hour of observation, this was significantly different (p<0.05) from the amphetamine treated group. The lowest dose 6.25 mg also significantly (p<0.05) reduced this index within 15-20 minutes of test time (figure 2).

Lophira alata and Afzelia africana significantly reduced sniffing and rearing in both models at some of the doses used for this study. Pretreatment with Lophira alata and Afzelia africana at doses of 6.25, 12.5 and 25 mg/kg showed a reduction in episodes of rearing while 12.5 and 25 mg/kg dose level significantly reduced constant rearing compared to amphetamine treated animals (Table 2).

Post treatment with *Lophira alata* and *Afzelia africana* significantly reduced episodes of rearing at all 3 dose levels used (Table 3).

Phytoconstituents	Lophira alata	Afzelia africana
Alkaloids	+	+
Reducing sugar	-	-
Saponins	+	+
Glycosides	+	+
Phytosterols	+	+
Tannis	+	+
Flavonoids	+	+
Triterpenoids	+	+

Table 1: Result of phytochemical screening of the aqueous extract of Lophira alata and Afzelia africana stem barks.

"+" represent class of phytochemicals present



Figure 1: Effect of pretreatment with *Lophira alata* and *Afzelia africana* on amphetamine induced locomotive activity Data are expressed as mean  $\pm$  SEM. \*p<0.05 compared to haloperidol, #p<0.05 compared to dextroamphetamine, n=5 per group. Dex indicates dextroamphetamine; AA, *Afzelia africana*; LA, *Lophira alata* 



Figure 2: Effect of post treatment with *Lophira alata* and *Afzelia africana* on Amphetamine induced locomotive activity Data are expressed as mean ± SEM. \*p<0.05 compared to haloperidol, #p<0.05 compared to dextroamphetamine, n=5 per group Dex indicates dextroamphetamine; AA, *Afzelia africana*; LA, *Lophira alata*.

Table 2: Effect of	pretreatment with La	ophira alata and a	Afzelia africana on J	Amphetamine induced stereotyp	)V

	Dex	6.25mg LA&AA + Dex	12.5mg LA&AA +	25mg LA&AA +	Haloperidol 1mg + Dex
		-	Dex	Dex	
IS	$0.28 \pm 0.13$	$0.52 \pm 0.12$	$0.53 \pm 0.13$	$0.65 \pm 0.13^*$	$0.00 \pm 0.00$
CS	$1.84 \pm 0.28^*$	$1.27 \pm 0.23^*$	$1.40 \pm 0.23^*$	$1.26 \pm 0.23$	$0.00 \pm 0.00^{\#}$
OR	$0.44 \pm 0.15$	$0.67 \pm 0.12$	$0.67 \pm 0.12$	$0.67 \pm 0.11$	$0.00 \pm 0.00$
RM	$1.06 \pm 0.29^*$	$0.57 \pm 0.19^{\#}$	$0.86 \pm 0.21^{*}$	$0.70 \pm 0.20^{\#}$	$0.00 \pm 0.00^{\#}$
RA	$0.82 \pm 0.37^*$	$0.92 \pm 0.28^*$	$0.10 \pm 0.11^{\#}$	$0.45 \pm 0.21^{*\#}$	$0.00 \pm 0.00^{\#}$

Data are expressed as mean  $\pm$  SEM. \*p<0.05 compared to haloperidol, #p<0.05 compared to dextroamphetamine, n=5 per group Dex indicates dextroamphetamine; AA, *Afzelia africana*; LA, *Lophira alata*; IS, intermittent sniffing; CS constant sniffing; OR occasional rearing; RM rearing most of the time; RA rearing all the time.

	Dex	Dex + 6.25mg LA&AA	Dex + 12.5mg	Dex + 25mg	Dex + Haloperidol 1mg
			LA&AA	LA&AA	
IS	$0.29 \pm 0.13$	$0.00 \pm 0.00$	$0.09 \pm 0.09$	$0.00 \pm 0.00$	$0.00 \pm 0.00^{\#}$
CS	$1.99 \pm 0.28^{*}$	$1.62 \pm 0.32^*$	$1.29 \pm 0.28^*$	$1.58 \pm 0.26^{*}$	$0.00 \pm 0.00^{\#}$
OR	$0.63 \pm 0.15^*$	$0.25 \pm 0.14$	$0.26 \pm 0.11$	$0.17 \pm 0.09^{\#}$	$0.00 \pm 0.00^{\#}$
RM	$1.33 \pm 0.29^*$	$0.82 \pm 0.29^*$	$0.26 \pm 0.15^{\#}$	$0.72 \pm 0.22^{*\#}$	$0.00 \pm 0.00^{\#}$
RA	$1.02 \pm 0.37^*$	0.96 ±0.37	$1.39 \pm 0.34^*$	$0.63 \pm 0.26^{\#}$	$0.00 \pm 0.00^{\#}$

Data are expressed as mean  $\pm$  SEM. \*p<0.05 compared to haloperidol, #p<0.05 compared to dextroamphetamine, n=5 per group Dex indicates dextroamphetamine; AA, *Afzelia africana*; LA; *Lophira alata*; IS, intermittent sniffing; CS constant sniffing; OR occasional rearing; RM rearing most of the time; RA rearing all the time.

#### DISCUSSION

This study was carried out to evaluate the antipsychotic potential of the aqueous extract of *Lophira alata and Afzelia africana* stem barks.

The preliminary phytochemical screening of aqueous extract of Lophira alata and Afzelia africana stem barks revealed the presence of alkaloids, tannins, saponins, phytosterols, flavonoids, triterpenoids and glycosides. Alkaloids, flavonoids, saponins, triterpenes and tannins have been reported to possess central nervous system modifying activity such as sedation, anxiolysis, psychotropic, depression, analgesic and anti-convulsant activity (Bhatachanya et al., 1997; Verma et al., 2010; Pritam et al., 2011).

The median lethal dose of aqueous extract of *Lophira alata* and *Afzelia africana* was estimated to be greater than 5000 mg/kg using the method described by Lorke (1983). This high median lethal dose (LD<sub>50</sub>) value suggests they are relatively non-toxic. The highest dose used in the study was less than 30% of the LD<sub>50</sub> which has been reported to be safe for use in ethnopharmacological assays (Matsumara, 1988; Vongtau *et al.*, 2004)

Amphetamine-induced stereotypy which measures dopamine  $D_2$  receptor reactivity mediated by hyperactivity of dopaminergic mechanisms in the nigrostriatal and mesolimbic pathway is an established animal model for schizophrenia (Valame and Gupata, 1981). Amphetamine, an indirectly acting sympathomimetic agent, releases catecholamines from neuronal storage pools and induces characteristic stereotyped behaviour in rats (Vogel, 2002; Kasture, 2006).

In this study, pre-treatment with a mixture of *Lophira alata* and *Afzelia africana* caused an initial reduction of hyper locomotion but increased locomotive activity which worsened with time. A distinct pattern of sleep was also observed in these animals. This effect could be due to the sedative potential of both extracts and probably displacement by amphetamine. The extracts could also be short-acting.

In order to simulate clinical psychosis, some animals were treated with amphetamine prior to treatment with graded doses of *Lophira alata* and *Afzelia africana*. Treatment with *Lophira alata* and *Afzelia africana* following amphetamine resulted in reduction of locomotion at a dose level of 12.5 mg/kg. At higher doses-50 and 100 mg/kg (data not shown), stereotypy was worsened; hyper locomotion in some animals could not be quantified due to very rapid movements while others exhibited rapid backwards and circular movements. These lasted for about four hours after which animals slept off and could not be roused for another 18 hours. We speculate that higher doses potentiate the effects of amphetamine though more mechanistic studies will be required to substantiate this hypothesis.

Sniffing and rearing are also indices of stereotypy and reduction of these indices are suggestive of sedative and/or antipsychotic activity (Costall and Naylor, 1973; Creese and Iverson, 1973; Costall and Naylor, 1974). In this study, different dose levels of *Lophira alata* and *Afzelia africana* reduced episodes of sniffing and rearing. Reduction in sniffing and rearing taken together with reduction in locomotor activity lends credence to the antipsychotic potential of *Lophira alata* and *Afzelia africana*.

#### CONCLUSION

Aqueous extract of *Lophira alata* and *Afzelia africana* stem bark extract possess anti psychotic properties which may account for their use as a mixture in ethnomedicine for management of psychosis.

#### REFERENCES

Abi-Dargham A, Laruelle M (2005). Mechanisms of action of second generation antipsychotic drugs in schizophrenia: insights from brain imaging studies. *Eur J Psychiatry* 20:15-27.

Adjanohoun E, Adjakidje V, Ahyi MRA, Akpagana K, Chibon P, El-Hadji A, GuinkoS, Siamevi KM. (1986). Contribution aux etudes ethnobotaniques et

floristiques au Togo. Agence de cooperation culturelle et technique (A.C.C.T), Paris pp.671.

Ahmed N, Azam NK (2014). Traditional knowledge and formulations of medicinal plants used by the traditional medical practitioners of Bangladesh to treat schizophrenia like psychosis. *Schizophrenia Res Treatment*. doi.org/10.1155/2014/679810

Ajiboye TO, Yakubu MT, Olajide AT (2014). Cytotoxic, anti-mutagenic and antioxidant activities of methanolic extract and chalcones dimers (lophirones B and C) derived from *Lophira alata* stem bark (Van Tiegh. Ex Keay). *J Evid Based CAM* 19:20-30.

Akah PA, Okpi O, Okoli CO (2007). Evaluation of the anti-inflammatory and analgesic activities of *Afzelia africana*. *Nig J Nat Prod Med* 11:48-52.

Akinpelu DA, Aiyegoro OA, Okoh AI. (2008). *In vitro* antimicrobial and phytochemical properties of crude extract of stem bark of *Afzelia africana* (Smith). *Afr J Biotech* 7:3665-3670.

Ayoka AO, Akomolafe RO, Iwalewa EO, Akanmu MA, Ukponmwan OE (2006). Sedative, anti-epileptic and antipsychotic effects of *Spondis mombin* L. (Anacardiaceae) in mice and rats. *J Ethnopahrmacol* 103:166-75.

Balde NM, Kaké A, Diallo MM, Balde MA, Maugendre D (2006). Herbal medicine and treatment of diabetes in Africa: an example from Guinea. *Diabetes Metab* 32:171-175.

Baldé AM, Traoré MS, Diallo MST, Baldé ES, Huang Y, Liu Z, Oularé K, Barry MS, Baldé MA, Camara A, Berghe VD, Vlietinck A, Pieters L (2015). Ethnobotanical survey, antimicrobial and anti-complement activities of Guinean medicinal plants traditionally used in the treatment of inflammatory diseases in Conakry and Dubreka. *J Plant Sci* 3:11-19.

Bhatacharya SK, Satyan KS. Bhatacharya SK, Satyan KS (1997). Experimental methods for evaluation of psychotropic agents in rodents:I-Anti-anxiety agents. *Indian J Exp Biol.* 35:565-575.

Bolza E, Keating WG (1972). African timbers: the properties, uses and characteristics of 700 species. Division of Building Research, CSIRO, Melbourne, Australia. pp 710.

Burkill HM Editor (1985). The useful plants of West Tropical Africa 2nd ed. London: Royal Botanical Gardens, Kew, Richmond, United Kingdom. pp 969.

Burt DR, Creese I, Snyder SH. (1977). Antischizophrenic drugs: chronic treatment elevates dopamine receptor binding in brain. *Science* 196:326-328.

Costall B, Naylor RJ (1973). The role of telencephalic dopaminergic systems in the mediation of apomorphine-stereotyped behaviour. *Eur J Pharmacol* 24:8-24.

Costall B, Naylor RJ (1974). Extrapyramidal and mesolimbic involvement with the stereotypic activity of d- and l-amphetamine. *Eur J Pharmacol* 25:121-129.

Creese I, Iverson SD (1973). Blockage of amphetamine-induced motor stimulation and stereotypy in the adult rat following neonatal treatment with 6-hydroxydopamine. *Brain Res* 55: 369-382.

Falade MO, Akinboye DO, Gbotosho GO, Ajaiyeoba EO, Happi TC, Abiodun OO, Oduola AMJ (2014). *In Vitro* and *In Vivo* antimalarial activity of *Ficus thonningii* Blume (Moraceae) and *Lophira alata* Banks (Ochnaceae), identified from the Ethnomedicine of the Nigerian Middle Belt. *J Par Res* doi: 10.1155/2014/972853

Gill LS (1992) Ethnomedicinal uses of plants in Nigeria. Uniben Press, Benin-City, Nigeria pp 15-65.

Hodgins S (1992). Mental disorder, intellectual deficiency, and crime: evidence from a birth cohort. *Arch Gen Psy* 6(49):476-483.

Ibrahim JA, Muazzam I, Jegede IA, Kunle OF, Okogun JI (2007). Ethno-medicinal plants and methods used by Gwandara tribe of Sabo Wuse in Niger State, Nigeria, to treat mental illness. *Afr. J. Trad. CAM* 4 (2):211-218.

Iwu MM (1993). A handbook of African Medicinal Plants, CRC Press, Florida, USA. pp 169,182

Iwu MM, Wootton JC, Eds (2002). Ethnomedicine and drug discovery. Elsevier, Amsterdam, Netherlands. pp 23-27 Kasture SB (2006). A Handbook of Experiments in preclinical pharmacology. Career Publications, Nashik, India. pp 43-110.

Lorke D (1983). A new approach to acute toxicity testing. *Arch Toxicol*. 54:275-287.

Magaji MG, Anuka JA, Abdu-Aguye I, Yaro AH, Hussaini IM (2008). Behavioural effects of the methanolic root bark extract of *Securinega virosa* in rodents. *Afr J Trad CAM* 5:147-153.

Magassouba FB, Diallra A, Camara A, Keita S, Barry MS, Donzo M, Bangoura O, Pieters L (2007). Ethnobotanical survey and antibacterial activity of some plants used in Guinea traditional medicine. *J Ethnopharmacol* 114:44-53

Matsumara F (1998). Toxicology of insecticides 2<sup>nd</sup> ed. Plenum Press, New York, pp 588

Meltzer H (2009). Basic and clinical Pharmacology 11<sup>th</sup> edition McGraw Hill, Singapore pp 487-501.

Murray CJL, Lopez AD Eds (1996). The global burden of disease: a comprehensive assessment of mortality and disability from diseases, injuries, and risk factors in 1990 and projected to 2020. Harvard School of Public Health, Cambridge, Massachutes, USA.

Musa H, Adeyinka AT, Tayo AA, Taiye MA (2013). Phytochemical analysis and antibacterial activity of *Khaya grandifoliola* and *Lophira alata* against some selected clinical bacterial isolates. *Wudpecker J Med Plants.* 2(1):7-15.

National Institute for Health, Public Health Service on Humane Care and Use of Laboratory Animals. USA, 2002.

Onweluzo JC, Onuoha KC, Obanu ZA (1995). A comparative study of some functional properties of *Afzelia africana* and *Glycine max* flours. *Food Chem* 54:55-59.

Pieme CA, Dzoyem JP, Kechia FA, Etoa FX, Penlap V (2008). *In vitro* antimicrobial activity of extracts from some Camerounian medicinal plants. *J Biol Sci* 8:902-907.

Pritam SJ, Amol BB, JS Sanjay (2011). Analgesic activity of *Abelmoschus monihot* extracts. Int *J Pharmacol.* 7:716-720.

Rang HP, Dale MM, Ritter JM, Flower RJ, Henderson G (2007). Rang and Dale's Pharmacology, 5<sup>th</sup> edition, Churchill Livingstone, London. pp 553-560

Romeiras MM, Duarte MC, Indjai B, Catarino L (2012). Medicinal plants used to treat neurological disorders in West Africa: a case study with Guinea-Bissau Flora. *Am J Plant Sci.* 3(7):1028-1036.

Sofowora A (2008). Medicinal plants and Traditional Medicine in Africa. Spectrum Books Ibadan pp 37-56, 106-108.

Sonibare MA, Soladoye MO, Subuloye TO (2008). Ethnobotanical survey of anti-psychotic plants in Lagos and Ogun States of Nigeria. *Eur J Sci Res.* 19:634-643.

Sonibare MA, Lawal TO, Ayodeji OO (2011). Antimicrobial evaluation of plants used in the management of psychosis opportunistic infections. *Int J of Pharmacol* 7(4):492- 497.

Tih EA, Tih RG, Sondengam BL, Martin MT, Bodo B (1994). Minor bioflavonoids from *Lophira alata* leaves. *J Nat Prod* 57:971-977.

Trease G E, Evans MC Editors (1983). Textbook of Pharmacognosy, 12th ed. London: Balliere Tindall. pp 322-383.

Valame SP, Gupata KC (1981). Effect of clonidine on amphetamine-induced stereotype. *Indian J Pharmacol.* 13:203-204.

Verma A, Jana GK, Sen S, Chakraborty R, Sachan SA, Mishra A (2010). Pharmacological evaluation of *Saraca indica* leaves for central nervous system depressant activity in mice. *J Pharm Sci Res.* 2:338-243.

Vogel HG, Editor (2002). Drug discovery and evaluation, pharmacological assays, 2nd ed. Berlin Heidelberg: Springer-Verlag pp 535.

Vongtau HO, Abbah J, Ngazal IE, Kunle OF, Chindo BA, Otsapa PB, Gamaliel KS (2004). Antinociceptive and anti-inflammatory activities of the methanolic extract of *Pinanari polyandra* stem bark in rats and mice. *J Ethnopharmacol* 90:115-21.

WHO fact sheet 134, accessed 24<sup>th</sup> September 2015. http://www.who.int/mediacentre/factsheets/ 2003/fs134/en/print.html