

**Synthesis, Characterization and Antibacterial Activities of Cu (II) Complex of Benzylpenicillin**

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**ABSTRACT**

The sudden uprising of resistant human pathogens is a challenge in current antimicrobial therapy, therefore the need to encourage the development of novel metal-based drugs. The efficacy of the various organic therapeutic agents can often be enhanced upon coordination with a suitable metal ion. Cu (II) benzylpenicillin, [Cu(Bpen)] complex was synthesized by the reaction of benzylpenicillin with CuCl<sub>2</sub>.2H<sub>2</sub>O. The complex was characterized by melting point, solubility, colour, conductivity, elemental analysis, infrared, UV/Visible, <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopy. The spectral data showed that the ligand benzylpenicillin behaved as a pentadentate ligand coordinating to the metal ion through OH, C=O of amide, C-O of β-lactam, C=O of COOH and NH. A trigonal bipyramidal geometry has been proposed for [Cu(Bpen)]. The ligand and [Cu(Bpen)] complex have been screened for their *in vitro* antibacterial activity against four gram positive strains (*Staphylococcus aureus*, *Bacillus subtilis*, *Bacillus cereus* and *Enterococcus faecalis*) and four gram negative (*Escherichia coli*, *Enterobacter cloacae*, *Pneumonia aeruginosa* and *Campylobacter felus*). It was deduced that [Cu(Bpen)] complex has improved antibacterial activity than benzylpenicillin alone and it was evident that the overall potency of benzylpenicillin was enhanced on coordination with copper ion.

**Keywords:** benzylpenicillin, copper, trigonal bipyramidal, complex, antibacterial

**INTRODUCTION**

The emergence of resistant human pathogens is a major problem in current antibacterial therapy, therefore there's need to encourage efforts towards the development of novel drugs and transition metal complexes that have been found to be particularly useful in this matter (Witkop and Ramachandran, 1964). Coordination compounds are important classes in inorganic chemistry that have been widely studied nowadays and they have been known to play vital roles in the field of medicine since the discovery of cis-platin and carboplatin as anticancer agents (Shechter and Karlish, 1980; Thompson *et al.*, 1999). Cisplatin, one of the first inorganic complexes discovered, is the most efficient drug for the treatment of certain types of cancer; however, drug toxicity and resistance limit its utilization for a broader range of diseases. Throughout the years of scientific research copper (II) complexes have been found to possess various activities such as anti-ulcer (Bonham *et al.*, 2002), anti-moebic (Rakel, 2007), anti-diabetic (Davis, 2003), anti-convulsant (Rottkamp *et al.*, 2000), anti-inflammatory (Adelstein and Vallee, 1961; Harris *et al.*, 1998; Christen, 2000), antimicrobial (Strausak *et al.*, 2001) and anti-tumor (Schaefer and Gitlin, 1999). Various copper (II)

complexes have been reported with antimicrobial activities. For instance, Copper (II) complex of Kefzol was screened against two gram (+) and gram (-) bacterial strains. It exhibited a marked enhancement in Kefzol activity against all the tested bacterial strains (Chohan *et al.*, 2004). Six copper(II) complexes of 2-benzoylpyridine N(4)-cyclohexylthiosemicarbazone were tested against five types of bacteria and were found to have an elevation in their bacteria activity against *Bacillus Sp.*, *Vibrio cholerae*, *Staphylococcus aureus* and *Salmonella paratyphi* (Joseph *et al.*, 2004). Novel complex of Cu (II) was synthesized from cyclohexane-1,3-dione of the type [Cu (L<sup>2</sup>)<sub>2</sub>].2NO<sub>3</sub>.H<sub>2</sub>O (where L<sup>2</sup> = 2-[2-(3-nitrophenyl)hydrazono]cyclohexane-1,3-dione). It was screened against *Escherichia coli* ATCC25922, *Enterococcus faecalis* ATCC29212, *Staphylococcus aureus* ATCC25923, and *Salmonella typhimurium* CCM583 respectively and was observed to show average level antibacterial activity against the bacteria compared to ampicillin (Turan *et al.*, 2015). Pahontu *et al.* (2016) synthesized new Cu (II) complexes from 8-ethyl-2-hydroxytricyclo[3.3.1]non-13-one-thiosemicarbazone.

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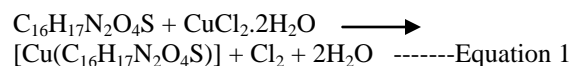
The complexes were tested for their antimicrobial activity against *Escherichia coli*, *Salmonella enteritidis*, *Staphylococcus aureus*, and *Enterococcus faecalis*. The result showed that all the complexes showed stronger antibacterial activity when compared to the free ligand. Common anti-bacterial agents have also been used as ligands to complex copper ions. It was noticed that the antimicrobial activity against *Mycobacterium smegmatis* of a metal ion complex in comparison to free ciprofloxacin, a bacterial gyrase inhibitor, increased three times (Patole *et al.*, 2003). This may have resulted from the facilitated diffusion of the drug through the cell membranes, presumably by an increase in the lipophilicity of the drug (Jimenez-Garido *et al.*, 2005). Complexes of cloxacillin with Co(II), Ni(II), Cu(II) and Zn(II) were synthesized and the coordination occurred through oxygen of carboxylate group and nitrogen of  $\beta$ -lactam group. These complexes were investigated and found to have an octahedral geometry with formula  $[M(\text{cloxa})(\text{H}_2\text{O})_3\text{Cl}]$  and they are non-electrolytic in nature. It was also found that the complexes have improved antibacterial activity than cloxacillin alone and it was evident that overall potency of cloxacillin was enhanced on coordination with metal ions (Chohan and Supuran, 2006). Bamigboye *et al.* (2012) synthesized three complexes of mixed sulfamethoxazole-cloxacillin drugs. Both the sulfamethoxazole and cloxacillin acted as a bidentate ligands towards Mn(II), Cu(II), Zn(II) metal ions and all the three complexes were assigned octahedral geometry. The coordination of metal ion with cloxacillin was through carbonyl oxygen and lactam ring oxygen, while the coordination of sulfamethoxazole was through sulphone oxygen and amine nitrogen group and to complete the coordination of octahedral geometry two chloride ions were intruded. Investigations of antimicrobial activities of the complexes against the tested microorganisms were found to be more active than their parent ligands. Thousands of coordination compounds have been literally prepared based on well-conceived ideas of improving their efficacy and have been subsequently screened over a wide range of diseases but few of them have been successful in passing the clinical tests (Kostova, 2010; Moues *et al.*, 2009; Rafique *et al.*, 2010). It is for this purpose that this study sought to extend the landscape of drug design. We therefore present the synthesis, characterization and antibacterial activities of Cu (II) complex of benzylpenicillin

## MATERIALS AND METHODS

All the chemicals used were of analytical grade. Benzylpenicillin was obtained from Shanxi Federal Pharmaceutical Company Limited, Shanxi, China. The melting points and decomposition temperature of benzylpenicillin and its Co(II) complex were determined using Gallenkamp melting point apparatus. The solubility of the ligand and the metal complex were tested using various solvents such as water, methanol, ethanol, n-Hexane, petroleum ether and dimethylsulfoxide (DMSO). The molar conductance of benzylpenicillin and its Cu(II) complex ( $10^{-3}$  M solution) were recorded using Jenway Conductivity Meter 4510. DMSO was used as the solvent. The conductivity of DMSO which was used as solvent was  $8.37 \text{ Ohm}^{-1}\text{mol}^{-1}\text{cm}^{-1}$ . The elemental analysis for C, N, H and S were obtained using a Perkin-Elmer 240B elemental analyzer. The UV-visible spectral measurement (190 – 900 nm) was obtained using UV-1800 series. The solvent used was Dimethylsulfoxide (DMSO). IR spectra were obtained on a Perkin Elmer Spectrum BX FT-IR spectrophotometer ( $4400\text{-}350 \text{ cm}^{-1}$ ) in KBr pellets. The NMR spectral measurement was recorded on nuclear magnetic resonance Bruker spectrophotometer at 400 MHz using tetramethylsilane as internal standard and DMSO-d<sub>6</sub> as solvent.

### Synthesis of benzylpenicillin - Cu(II) complex

The complex was prepared following reported procedure by Anacona and Figueroa, (1999). Exactly 2.216 g (0.013 mole)  $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ , in 10 ml of water was added to a solution of benzylpenicillin (4.35 g, 0.013 mole). The mixture was stirred for 1 hour and the solid complex which separated was removed by filtration and washed with water, ethanol and ether. The complex was dried under vacuum at room temperature for 48 hours. The complex was then stored in a neat and labelled container after determining their percentage yield. The general synthesis for the benzylpenicillin - Cu(II) complex is proposed in Equation 1.



### Antibacterial activity test

The organisms used were Gram-negative *Escherichia coli*, *Enterobacter cloacae*, *Pneumonia aeruginosa*, and *Campylobacter felus*. The Gram-positive bacterial strains were *Staphylococcus aureus*, *Bacillus substilis*, *Bacillus cereus*, and *Enterococcus faecalis*. These clinical isolates were obtained from Federal Medical Centre, Umuahia, Abia State. Antibacterial activity of samples were determined by

using agar well diffusion method and bacterial growth were subcultured on nutrient broth for their *in vitro* testing which were prepared by dissolving (24 g) of nutrient broth. The mixture was autoclaved for 15 minutes at 120 °C. Stock solution for *in vitro* antibacterial activity was prepared by dissolving 5 mg of compound in 9 mL of DMSO. Inoculation was done with the help of micropipette with sterilized tips and 100 µL of activated strain was placed onto the surface of agar plate. It was spread over the whole surface and then two wells having diameter of 10 mm

were dug in media and incubated at 37 °C for 48 hours. Activity was determined by measuring the diameter of zone showing complete inhibition and has been expressed in mm.

#### Statistical analysis

Statistical significance was determined using Duncan Multiple Range Test. Results were considered statistically significant at  $P < 0.05$  and were expressed as mean  $\pm$ SD.

## RESULTS

Table 1: Some physicochemical properties of Benzylpenicillin and its copper complex

| Ligand/complex | Colour | M.P.(°C) | Yield % | Conductance $\text{Sm}^2.\text{mol}^{-1}$ | C (%)<br>Found<br>(Calc.) | H (%)<br>Found<br>(Calc.) | N (%)<br>Found<br>(Calc.) | S (%)<br>Found<br>(Calc.) |
|----------------|--------|----------|---------|---|---------------------------|---------------------------|---------------------------|---------------------------|
| Bpen           | White  | 209      | -       | 236.0                                     | 53.92<br>(53.77)          | 4.81<br>(4.73)            | 7.87<br>(7.88)            | 8.98<br>(8.88)            |
| [Cu(Bpen)]     | Black  | 200      | 72      | 146.9                                     | 45.75<br>(48.54)          | 45.75<br>(48.54)          | 6.67<br>(7.08)            | 7.62<br>(8.10)            |

Bpen = Benzylpenicillin

Table 2: Solubility profile of benzylpenicillin and its copper complex in some selected solvents

| Ligands/Complex | n-Hexane | Distilled water | Ethanol | Methanol | Petroleum ether | DMSO |
|-----------------|----------|-----------------|---------|----------|-----------------|------|
| Bpen            | S        | S               | S       | S        | S               | S    |
| [Cu(Bpen)]      | IS       | IS              | SS      | SS       | IS              | S    |

Key: S-Soluble, SS-Slightly Soluble, IS-Insoluble; Bpen = Benzylpenicillin

Table 3: Infrared spectral data of benzylpenicillin and its copper complex

| Ligand/complex | OH of COOH | N-H     | C=O of lactam | C=O of amide | $V_{\text{assym}}$ (COO) | $V_{\text{sym}}$ (COO) | C-O     |
|----------------|------------|---------|---------------|--------------|--------------------------|------------------------|---------|
| Bpen           | 3542.26    | 3351.48 | 1778.04       | 1697.66      | 1620.54                  | 1418.40                | 1161.61 |
| [Cu(Bpen)]     | absent     | 3248.50 | Absent        | 1640.37      | 1449.73                  | 1362.26                | 1124.00 |

Bpen = Benzylpenicillin

Table 4: Summary of the UV/Vis peaks of benzylpenicillin and its copper complex

| Ligand/Complex | Chromophores | Transitions   | $\lambda_{\max}$ nm   |
|----------------|--------------|---------------|---|
| Bpen           | C=C          | $\pi - \pi^*$ | 197.50  |
|                | C=O          | $n - \pi^*$   | 203.50, 209.50, 215.50, 226.50, 238.50, 255.50, 259.50, 270.50, 278.50, 284.50 and 317.50 |
| [Cu(Bpen)]     | C=C          | $\pi - \pi^*$ | 197.50  |
|                | C=O          | $n - \pi^*$   | 203.50, 209.50, 215.50, 226.50, 238.50, 255.50, 259.50, 270.50, 278.50, 284.50 and 317.50 |
|                |              | LMCT          | 366.80  |

Bpen = Benzylpenicillin LMCT = Ligand to metal charge transfer

Table 5: Summary of  $^1\text{H}$  NMR peaks benzylpenicillin and its copper complex

| Compound   | O-H (ppm) | C=OCH of $\beta$ -lactam (ppm) | NH of amide (ppm) | Methyl protons (ppm) | Ar H (ppm) |
|------------|-----------|--------------------------------|-------------------|----------------------|------------|
| [Bpen]     | 11        | 5.35                           | 8.72              | 1.46<br>1.59         | 7.12-7.32  |
| [Cu(Bpen)] | absent    | absent                         | 7.70              | 1.55<br>1.29         | 7.12-7.45  |

Bpen = Benzylpenicillin

Table 6: Summary of the  $^{13}\text{C}$  NMR peaks benzylpenicillin and its copper complex

| Compounds  | Phenyl carbons (ppm) | C=O of $\beta$ -lactam (ppm) | C=O Amide (ppm) |
|------------|----------------------|------------------------------|-----------------|
| [Bpen]     | 126.88-129.54        | 173.73                       | 170.86          |
| [Cu(Bpen)] | 125.98-129.05        | 152.50                       | 153.68          |

Bpen=Benzylpenicillin

Table 7: Percentage zone of inhibition (mm) of the benzylpenicillin and its copper (II) complex against gram positive bacterial population

| Ligand/complex | <i>Staphylococcus aureus</i> | <i>Bacillus subtilis</i> | <i>Bacillus cereus</i> | <i>Enterococcus faecalis</i> |
|----------------|------------------------------|--------------------------|------------------------|------------------------------|
| Bpen           | 2.52±0.03 <sup>a</sup>       | 6.12±0.03 <sup>a</sup>   | 4.96±0.01 <sup>a</sup> | 2.12±0.03 <sup>a</sup>       |
| [Cu(Bpen)]     | 5.60±0.08 <sup>b</sup>       | 10.73±0.24 <sup>b</sup>  | 8.15±0.02 <sup>b</sup> | 5.26±0.08 <sup>b</sup>       |

Means with different superscript are significantly different from each other ( $P < 0.05$ )

Table 8: Percentage zone of inhibition (mm) of the benzylpenicillin and its copper(II) complex against gram negative bacterial population

| Ligand/complex | <i>Escherichia coli</i> | <i>Enterobacter cloacae</i> | <i>Pneumonia aeruginosa</i> | <i>Campylobacter felus</i> |
|----------------|-------------------------|-----------------------------|-----------------------------|----------------------------|
| Bpen           | 10.43±0.03 <sup>a</sup> | 1.32±0.03 <sup>a</sup>      | 6.82±0.03 <sup>a</sup>      | 7.32±0.02 <sup>a</sup>     |
| [Cu(Bpen)]     | 16.84±0.02 <sup>b</sup> | 8.11±0.01 <sup>b</sup>      | 14.82±0.05 <sup>b</sup>     | 10.44±0.01 <sup>b</sup>    |

Means with different superscript are significantly different from each other ( $P < 0.05$ )



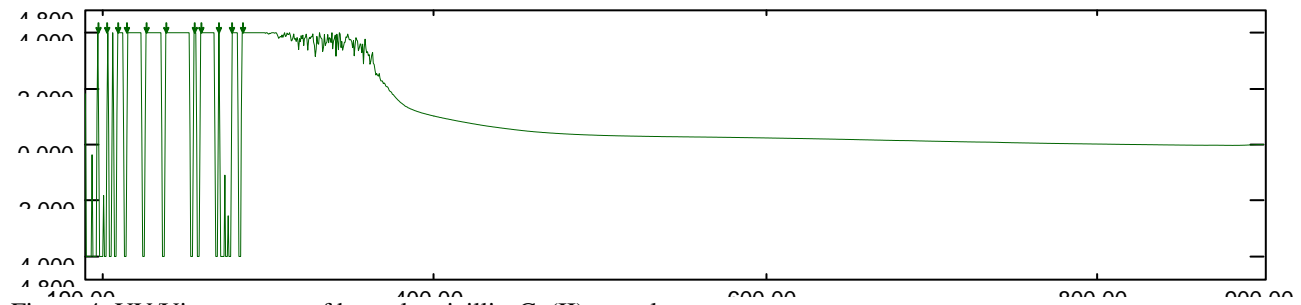


Figure 4: UV/Vis spectrum of benzylpenicillin-Cu(II) complex

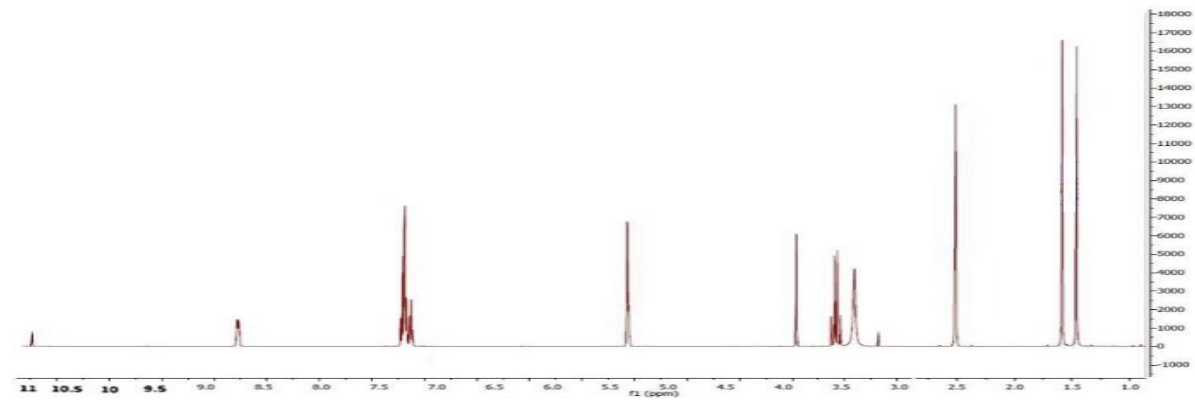


Figure 5: <sup>1</sup>H NMR Spectrum of Benzylpenicillin

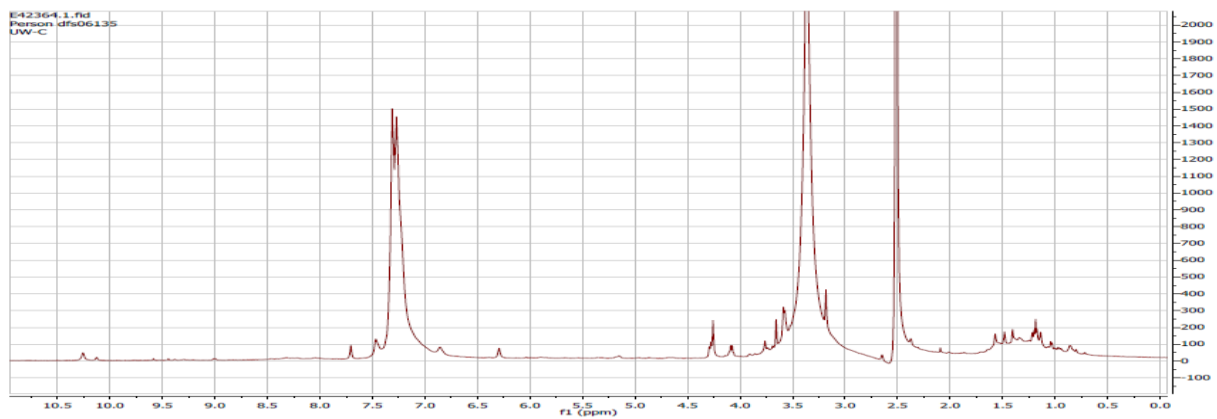


Figure 6: <sup>1</sup>H NMR Spectrum of benzylpenicillin-Cu(II) complex

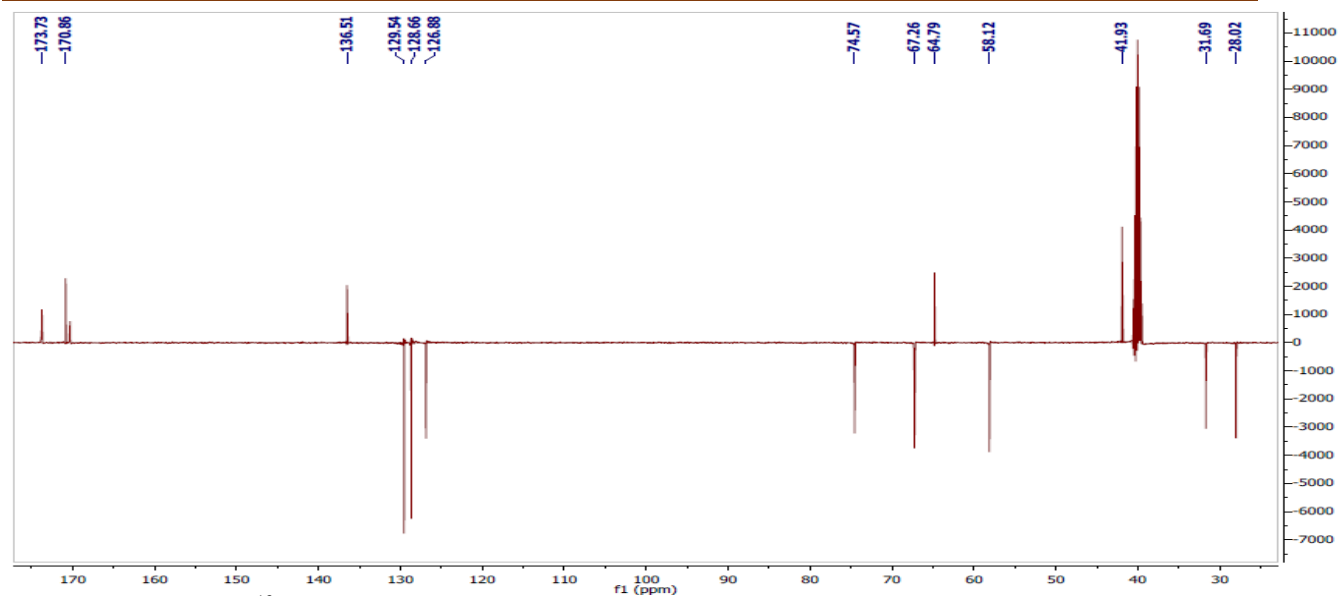


Figure 8:  $^{13}\text{C}$  NMR (DEPT 135) Spectrum of benzylpenicillin

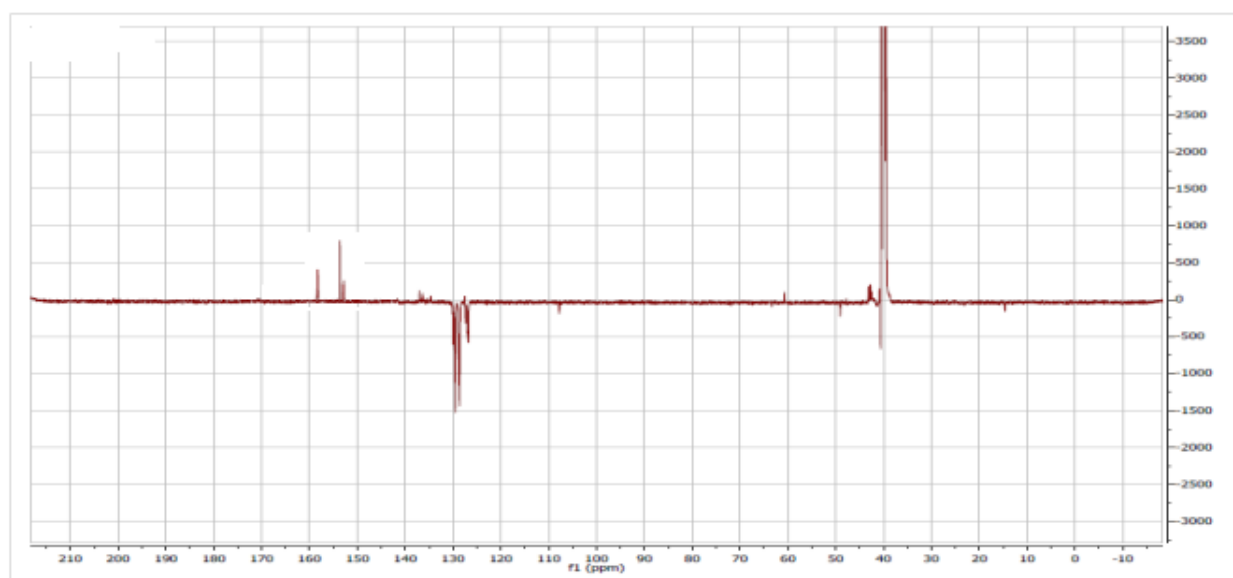


Figure 9:  $^{13}\text{C}$  NMR (DEPT 135) Spectrum of benzylpenicillin-Cu (II) complex

[Cu(Bpen)] is a non-hygroscopic, air and photo stable crystalline powder. It is black in colour with a melting of  $200\text{ }^{\circ}\text{C}$  lower than the parent antibiotic ( $209\text{ }^{\circ}\text{C}$ ) (Table 1). The percentage yield of 72 % was very good. The micro analytical measurements and metal estimation data results suggested 1:1 metal ligand ratio and formation of the complex. The conductivity measurements in DMSO (Table 1) revealed that the ligand and complex were electrolyte with a value  $236.0$  and  $146.9\text{ Sm}^2\text{mol}^{-1}$  (Geary,

1971). The conductivity of DMSO which was used as solvent was  $8.37\text{ Sm}^2\text{mol}^{-1}$ . The solubility of benzylpenicillin and [Cu(Bpen)] in various solvents is shown in Table 2. Benzylpenicillin was found to be soluble in distilled water, n-hexane, ethanol, methanol, petroleum ether and DMSO. The complex was found to be insoluble in distilled water, n-hexane and petroleum ether. It was slightly soluble in ethanol and methanol but completely soluble in DMSO. The solubility data suggested that the complex is

moderately polar. Infrared spectral data of benzylpenicillin and its copper complex are shown in Table 1 while infrared spectra are presented in Figure 1 and 2 respectively. The vibration frequency  $1697.66\text{ cm}^{-1}$  was assigned to C=O stretch of carboxylic acid. In the IR spectrum of [Cu(Bpen)] this wavenumber was shifted to  $1640.37\text{ cm}^{-1}$ . This suggests that coordination occurred through C=O of COOH.  $\beta$ -lactam carbonyl group which was observed at  $1778.04\text{ cm}^{-1}$  in the spectrum of benzylpenicillin was absent in the spectrum of [Cu(Bpen)]. This suggested the involvement of  $\beta$ -lactam carbonyl group in complex formation, hence, the formation of C-O $\rightarrow$ M bonding system. This also suggests that the  $\beta$ -lactam C=O was converted to C-O during complexation. The vibration frequency  $3542.26\text{ cm}^{-1}$  was assigned OH of COOH in the IR spectrum of benzylpenicillin. This vibration frequency was absent in the IR spectrum of [Cu(Bpen)]. This suggests that OH was deprotonated during coordination. In the spectrum of the ligand, the wavenumber  $3351.48\text{ cm}^{-1}$  was assigned N-H stretch. This wavenumber was shifted to a lower wavenumber ( $3248.50\text{ cm}^{-1}$ ) in the complex. This suggests that complexation occurred through N-H function group because increase in electron density, increases the N-H bond length and consequently slows down the vibration frequency. The electronic spectral data of benzylpenicillin and [Cu(Bpen)] is shown in Table 4. The spectra are presented in Figure 3 and 4. The multiplet signals observed in the ligand and complex in the range  $\delta = 7.12\text{--}7.45\text{ ppm}$  were assigned to aromatic protons. Finally, singlets observed at  $\delta = 1.46$  and  $1.59\text{ ppm}$  were attributed to the two methyl protons on the thiazolidine ring of benzylpenicillin were observed at  $\delta = 1.29$  and  $1.55\text{ ppm}$  in the spectrum of the metal complex.

The  $^{13}\text{C}$  NMR (DEPT 135) spectral data of benzylpenicillin and [Cu(Bpen)] is shown in Table 6. The  $^{13}\text{C}$  NMR (DEPT 135) spectra are presented in Figures 8 and 9. In the  $^{13}\text{C}$  NMR spectrum of benzylpenicillin, C=O of amide was observed at  $170.86\text{ ppm}$ . The band for C=O of amide was shifted upfield  $153.68\text{ ppm}$  in the  $^{13}\text{C}$  NMR spectrum of [Cu(Bpen)]. This shift suggested that C=O of amide was involved in complexation to Cu(II) ion. C=O of carboxylic acid was observed at  $173.73\text{ ppm}$  in the  $^{13}\text{C}$  NMR spectrum of the ligand. This band was shifted upfield ( $152.50\text{ ppm}$ ) in the  $^{13}\text{C}$  NMR spectrum of the complex. This also suggested that C=O of carboxylic acid was involved in complexation to Cu(II) ion. The total numbers of carbons were equal to the peaks in the DEPT experiment.

Figures 3 and 4. The electronic spectrum of benzylpenicillin maximally absorbed at  $\lambda = 197.50, 203.50, 209.50, 215.50, 226.50, 238.50, 255.50, 259.50, 270.50, 278.50, 284.50$  and  $317.50\text{ nm}$  and these absorptions were as a result of the chromophores present in benzylpenicillin. These transitions have been assigned  $\pi - \pi^*$  and  $n - \pi^*$  and are known as Intra-ligand Charge Transfer (ILCT). The electronic spectrum of [Cu(Bpen)] maximally absorbed at  $\lambda = 197.50, 203.50, 209.50, 215.50, 226.50, 238.50, 255.50, 259.50, 270.50, 278.50, 284.50$  and  $317.50\text{ nm}$ . These transitions have been assigned  $\pi - \pi^*$  and  $n - \pi^*$  and are known as Intra-ligand Charge Transfer (ILCT). The absorption band at  $\lambda = 561.00\text{ nm}$  in the spectrum of the complex have been assigned ligand to metal charge transfer (LMCT). The  $^1\text{H}$  NMR spectral data of benzylpenicillin and [Cu(Bpen)] are shown in Table 5. The  $^1\text{H}$  NMR spectra are presented in Figures 4 and 5. The singlet at  $11.00\text{ ppm}$  corresponded to hydroxyl group of carboxylic acid. This chemical shift was absent in the  $^1\text{H}$  NMR spectrum of [Cu(Bpen)]. This suggested that  $-\text{OH}$  was deprotonated during coordination. The doublet observed at  $\delta = 8.72\text{ ppm}$  in  $^1\text{H}$  NMR of benzylpenicillin was assigned to NH proton of amide. This chemical shift was shifted to  $7.70\text{ ppm}$  in the  $^1\text{H}$  NMR spectrum of [Cu(Bpen)]. This shift suggested that coordination occurred through N-H functional group.

Based on elemental analysis, IR, UV-Visible,  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR, the structure in Figure 10 have been proposed for benzylpenicillin -copper (II) complex. The structure of benzylpenicillin is shown in Figure 11.

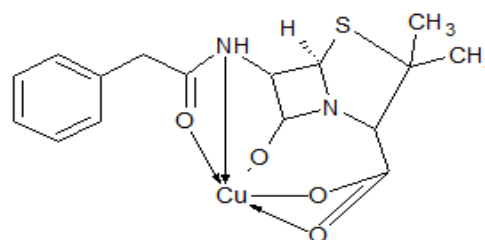


Figure 10: Suggested structure of benzylpenicillin -copper (II) complex

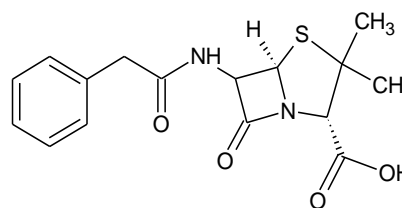


Figure 11: Structure of benzylpenicillin



The antibacterial activity of benzylpenicillin and its copper (II) complex against four gram positive bacterial strains (*Staphylococcus aureus*, *Bacillus subtilis*, *Bacillus cereus*, and *Enterococcus faecalis*) is presented in Table 7 while the antibacterial activity of benzylpenicillin and its copper (II) complex against four gram negative bacterial strains (*Escherichia coli*, *Enterobacter cloacae*, *Pneumonia aeruginosa*, and *Campylobacter felus*) is presented in Table 8. The result showed that the synthesized compound exhibited varying degree of inhibitory results on the growth of different bacterial strains. The inhibitory activity of the complex [Cu(Bpen)] was significantly higher ( $p < 0.05$ ) against *Staphylococcus aureus*, *Bacillus subtilis*, *Bacillus cereus*, *Enterococcus faecalis*, *Campylobacter felus*,

## REFERENCES

Adelstein, S.J. and Vallee, B.L. (1961). Copper metabolism in man. *New England Journal of Medicine*. 265: 892-897.

Anacona, J.R., and Figueroa, E.M. (1999). Synthesis and characterization of metal complexes with penicillin. *Journal of Coordination Chemistry*. 48(2): 181-189.

Bamigboye, M.O., Obaleye, J.A. and Abdulmolib, S. (2012). Synthesis, characterization and antimicrobial activity of some mixed sulfamethaxazole-cloxacillin metal drug complexes. *International Journal of Chemistry*. 22(2): 105-108.

Bonham, M., Jacqueline, M., Bernadette, M.H and Strain, J.J. (2002). The immune system as a physiological indicator of marginal copper status? *British Journal of Nutrition* 87: 393-403.

Chohan, Z.H. and Supuran, C.T. (2006). Metalloantibiotics: Synthesis, characterization and in-vitro antibacterial studies on cobalt (II), copper (II), nickel (II) and zinc (II) complexes with cloxacillin. *Journal of Enzyme Inhibition Medicinal Chemistry*. 21(4): 441 - 44.

Chohan, Z.H., Supuran, C.T. and Scozzafara, A. (2004). Metalloantibiotics: Synthesis and Antibacterial Activity of Cobalt(II), Copper(II), Nickel(II) and Zinc(II) complexes of Kefzol. *Journal of Enzyme Inhibition and Medicinal Chemistry*. 19(1): 79-84.

*Enterobacter*, *Pneumonia aeruginosa* and *Escherichia colias* compared to benzylpenicillin. This may have been as a result of the facilitated diffusion of the drug through the cell membranes, presumably by increasing the lipophilicity of the drug (Jimenez-Garido et al., 2005).

## CONCLUSION

Copper(II) complex of benzylpenicillin was synthesized. Benzylpenicillin and its copper (II) complex were characterized and tested for their antibacterial activity. The benzylpenicillin ligand was found to behave as a pentadentate ligand and a trigonal bipyramidal geometry was suggested for the metal complex. The complex was also found to have improved antibacterial activity than the free ligand.

Christen, Y. (2000). Oxidative stress and Alzheimer's disease. *American Journal of Clinical Nutrition*. 71: 621-629.

Davis, C.D. (2003). Low dietary copper increases fecal free radical production, fecal water alkaline phosphatase activity and cytotoxicity in healthy men. *Journal of Nutrition*. 133(2): 522-527.

Geary, W.J. (1971). The use of conductivity measurements in organic solvents for the characterization of coordination compounds. *Coordination Chemistry Reviews*. 7(1): 81-122.

Harris, E.D., Qian, Y., Tiffany-Castiglioni, E., Lacy, A.R. and Reddy, M.C. (1998). Functional analysis of copper homeostasis in cell culture models: a new perspective on internal copper transport. *American Journal of Clinical Nutrition*. 67: 988-995.

Jimenez-Garido, N, Perello, L., Ortiz, R., Alzuet, G., Gonzalez-Alvarez, M., Canton, E., Liu Gonzalez, M., Garcia-Granda, S. and Perez-Priede, M. (2005). Antibacterial studies, DNA oxidative cleavage, and crystal structures of Cu(II) and Co(II) complexes with two quinolone family members, ciprofloxacin and enoxacin. *Journal of Inorganic Biochemistry*. 99: 677-689.

Joseph, M., Suni, V., Kurup, M.R., Nethaji, M., Kishore, A. and Bhat, S. (2004). Structural, spectral and antimicrobial studies of copper (II) complexes of 2-benzoylpyridineN(4)-cyclohexylthiosemicarbazone. *Polyhedron*. 23(18): 3069-3080.

Kostova, I. (2010). Metal-containing drugs and novel coordination complexes in therapeutic anticancer applications-part II. *Anticancer Agents in Medicinal Chemistry*. 10: 352-353.

Moues, C.M., Heule, F., Legerstee, R. and Hovius, R.N. (2009). Topical Negative Pressure in Wound Care Effectiveness and guidelines for clinical application. *Ostomy and wound management*. 55: 16-32.

Pahontu, E., Paraschivescu, C., Poirier, D., Oprean, C., Păunescu, V., Gulea, A., Roșu, T. and Bratu, O (2016). Synthesis and characterization of Novel Cu(II), Pd(II) and Pt(II) complexes with 8-Ethyl-2-hydroxytricyclo(7.3.1.0<sup>2,7</sup>)tridecan-13-one-thiosemicarbazone: Antimicrobial and *in vitro* Antiproliferative Activity. *Molecules*. 21(5): 674-678

Patole, J., Sandbhor, U., Subhash, P., Deobagkar, D.N. Anson, C.E. and Powell, A (2003). Structural chemistry and *in vitro* antitubercular activity of acetylpyridine benzoyl hydrazone and its copper complex against *Mycobacterium smegmatis*. *Bioorganic Medicinal Chemistry Letters*. 13: 51-55.

Rafique, S., Idrees, M. Nasira, A. Akbar, H. and Atha, A. (2010). Transition Metal Complexes as Potential Therapeutic Agents. *Biotechnological and Molecular Biological review*. 5(2): 38-45.

Rakel, D. (2007). Integrative Medicine (2nd edition), Elsevier. *The Journal of science and healing*. 3(5): 543-544.

Rottkamp, C.A., Nunomura, A., Raina, A.K., Sayre, L.M., Perry, G. and Smith, M. (2000). Oxidative stress, antioxidants, and Alzheimer's disease. *Alzheimer Disease Associated Disorders*. 14: 62-66.

Schaefer, M. and Gitlin, J.D. (1999). Genetic disorders of membrane transport IV-Wilson's disease and Menkes disease. *American Journal of Physiology*, 311 - 314

Shechter, Y. and Karlsh, S.D.J. (1980). Insulin-like stimulation of glucose oxidation in rat adipocytes by Vanadyl (IV) ions. *Nature*. 284: 556-558.

Strausak, D., Mercer, J.F., Dieter, H.H., Stremmel, W. and Multhaup, G. (2001). Copper in disorders with neurological symptoms: Alzheimer's, Menkes, and Wilson diseases. *Brain Research Bulletin* 55(2): 175-185.

Thompson, K.H., McNeill, J.H. and Orvig, C. (1999). Vanadium compounds as insulin mimics, *Chemical Reviews*. 99: 2561 – 2571.

Turan, N., Korkoca, H., Adigüzel, R., Çolak, N. and Buldurun, K. (2015). Synthesis, structural characterization and biological activity of novel cyclohexane-1,3-dione ligands and their metal complexes. *Molecules* 20(5): 9309-9325.

Witkop, B and Ramachandran, L.K. (1964). Progress in Non-enzymatic Selective Modification and Cleavage of Proteins. *Metabolism* 13(10): 1016-1025.